Hindawi Publishing Corporation Case Reports in Infectious Diseases Volume 2012, Article ID 453685, 3 pages doi:10.1155/2012/453685

# Case Report

# Necrotizing Fasciitis Associated with Staphylococcus lugdunensis

## Tony Hung,<sup>1,2</sup> Soroush Zaghi,<sup>1</sup> Jonathan Yousefzadeh,<sup>1</sup> and Matthew Leibowitz<sup>2</sup>

- <sup>1</sup> Department of General Surgery, David Geffen School of Medicine at UCLA, Los Angeles, CA 90095, USA
- <sup>2</sup> Division of Infectious Diseases, Department of Medicine, David Geffen School of Medicine at UCLA, Los Angeles, CA 90095, USA

Correspondence should be addressed to Soroush Zaghi, soroush.zaghi@gmail.com

Received 5 February 2012; Accepted 21 March 2012

Academic Editors: A. Marangoni and X. Vallès

Copyright © 2012 Tony Hung et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Necrotizing fasciitis is a life-threatening soft tissue infection that results in rapid local tissue destruction. Type 1 necrotizing fasciitis is characterized by polymicrobial, synergistic infections that are caused by non-Group A *streptococci*, aerobic and anaerobic organisms. Type 2 necrotizing fasciitis involves Group A *Streptococcus* (GAS) with or without a coexisting staphylococcal infection. Here we provide the first report of necrotizing fasciitis jointly associated with the microbes Group B *Streptococcus* and *Staphylococcus lugdunensis*. *S. lugdunensis* is a commensal human skin bacterium known to cause often painful and prolonged skin and soft tissue infections. To our knowledge, however, this is the first case of *Staph. lugdunensis*-associated necrotizing fasciitis to be reported in the literature.

#### 1. Introduction

The spectrum of disease-causing microbes continues to change rapidly. Not only are we documenting an alarming increase in antimicrobial resistance but also we are now detecting an emerging tendency for infectious disease patho gens to develop greater degrees of virulence. Necrotizing fasciitis (NF) is a rapidly progressing, life- and limbthreatening soft tissue infection that targets the superficial fascial tissue layer. The disease is characterized by quickly spreading erythema, pronounced tenderness, severe pain, subcutaneous gas, fever, and tachycardia [1]. Whereas a number of pathogens including Group A Streptococcus, Enterobacteriaceae, Clostridium perfringens, and Staphylococcus aureus have commonly been associated with necrotizing fasciitis, a growing number of unprecedented bacteria are also becoming associated with this severe soft tissue infection. As necrotizing fasciitis is a potentially lethal and devastating infectious disease process, it is important that novel virulence patterns be documented and addressed as soon as they are detected. Here we report the first case of necrotizing fasciitis to be associated with the microbe Staphylococcus lugdunensis.

#### 2. Case

A 66-year-old African American woman with osteoporosis, osteoarthritis, and impaired glucose tolerance presented to the emergency department with a 5-day history of left groin pain, nausea, vomiting, and fatigue. Five days prior to admission, she had noted an abscess on her inner left thigh that progressively ruptured and drained purulent, bloody material. On evaluation, she was tachycardic (HR: 111-136) and febrile to 103.2°F. The left thigh wound drained a thin serous and dishwater-type fluid. Labs revealed leukocytosis and hyperglycemia (WBCs =  $23.8 \times 10^3$ : 86.4% neutrophils, 5.4% lymph, 8.1% monocyte; glucose: 239 g/dL; BUN 13 mg/dL, creatinine 0.6 mg/dL, chloride 92 mmol/L, total CO<sub>2</sub> 24 mmol/dL, total creatinine kinase = 67 mg/dL). CT scan showed a  $4.6 \times 1.4 \times 5$  cm fluid collection proximal to the left gracilis muscles with presence of gas bubbles, edema, and fat stranding in the subcutaneous tissues. The patient was taken emergently to the operating room to undergo radical debridement, washout, packing, and subsequently admitted for IV antibiotic therapy.

Microscopic analysis of the debrided specimen revealed areas of necrosis and acute and chronic inflammation,

```
07/15/10 1700
                 (COLLECTION TIME)
                                       07/15/10 1733 (IN LAB TIME)
                                       FINAL ON 07/21/10 1517
-S BACTERIAL CULTURE
    SPECIMEN TYPE
                                       WOUND
    SOURCE-BODY SITE
                                       GROIN
    ORGANISM 1
                                       BETA STREPTOCOCCUS GROUP B
     OUANTITATION
    ORGANISM 2
                                       STAPHYLOCOCCUS LUGDUNENSIS
     QUANTITATION
                                       MODERATE
     SENSITIVITY SET-UP DATE:
                                       07/20/10
    ORGANISM 3
                                       CORYNEBACTERIUM SPECIES
     QUANTITATION
                                       MODERATE
    MIC in mcg/mL
                                      COST/
                                                ORG 2
_
        *USUAL ADULT IV DOSAGE
                                      DAY*
_
     CLINDAMYCIN
                                      $ 24
                                                 <=0.5 S
     DOXYCYCLINE
                                      $ 32
                                                  <=1 S
                                                 <=0.5 S
                                      $ 35
     ERYTHROMYCIN
     OXACILLIN
                                      $ 70
                                                   0.5 S
     PENICILLIN
                                      $ 38
                                                  0.06 S
                                      $ 33
                                                 <=0.5 S
     RIFAMPIN
     VANCOMYCIN
                                      $ 21
                                                     1 S
                  Erythromycin Susceptible STAPHYLOCOCCUS LUGDUNENSIS
                  are susceptible to azithromycin and clarithromycin.
                  Oxacillin-susceptible staphylococci are
                  susceptible to cefazolin, cephalexin and ceftriaxone.
```

Figure 1: Microbiology culture and sensitivity report from initial debridement.

consistent with a diagnosis of necrotizing fasciitis. Gram stain of the initial intraoperative specimen showed "few gram positive cocci in pairs and chains," and final bacterial culture showed many GBS, moderate S. lugdunensis, and moderate Corynebacterium (see Figure 1). Subsequent additional debridement grew many GBS and many S. lugdunensis. Blood cultures were negative. Blood tests for HIV 1, HIV 2, hepatitis B, and hepatitis C were negative. QuantiFERON gold tuberculosis test was negative. During the course of hospitalization, the patient was empirically treated with IV antibiotics including vancomycin, clindamycin, and aztreonam. After multiple debridements, the patient underwent plastic surgery reconstruction of her left groin. She was discharged home in stable condition after a 67-day hospital course and is doing well at followup 18 months later.

### 3. Discussion

Necrotizing fasciitis (NF) is a life-threatening soft tissue infection that results in rapid local tissue destruction of primarily the superficial fascia; the infective process is characterized by angiothrombotic microbial invasion and liquefactive necrosis of the deep dermis and fascia [1]. Estimated 500 to 1,500 cases of NF are diagnosed each year in the United States with a case-fatality rate of 24% [2]. While surgical debridement is the essential treatment of NF, addition of broad-spectrum antibiotics at the time of presentation and further appropriate antimicrobial therapy targeted for specific causative microbes may modify the

clinical picture and are also critical in the management of this infectious process [1, 2].

NF is traditionally classified into two distinct categories based on the causative microorganisms [2]. Type 1 NF is characterized by polymicrobial, synergistic infections that are caused by non-Group-A streptococci, aerobic and anaerobic organisms [1]. Type 2 NF involves Group A *Streptococcus* (GAS) with or without a coexisting staphylococcal infection [2]. While the clinical manifestations are similar, type 1 and type 2 NF tend to affect different subgroups of patients. Type 1 NF has a propensity to affect diabetic and immunocompromised individuals, whereas type 2 NF tends to occur in individuals with no underlying comorbidities [1]. Both type 1 and type 2 NF are considered surgical emergencies; appropriate management requires comprehensive surgical debridement combined with targeted antimicrobial therapy and physiologic support [3].

Because of the importance of targeted antibiotic therapy in the treatment of NF, awareness of the spectrum of microbes that might be associated with NF is imperative. Microbes commonly associated with NF include: Group A Streptococcus, Clostridium perfringens, methicillin-sensitive Staphylococcus aureus, Enterobacteriaceae, E. coli, Pseudomonas spp., Klebsiella spp., V. vulnificus, A. hydrophila, and anaerobic Streptococcus spp. [1]. While these microbes comprise most of all cases, other pathogens such as MRSA and Group B Streptococcus (GBS) have also been associated with NF on rare occasions [4]. Here we document for the first time a case of NF caused by a polymicrobial combination of Group B Streptococcus and Staphylococcus lugdunensis.

Staphylococcus lugdunensis is a commensal human skin bacterium first described in 1988 and since reported as a cause of prolonged and recurrent skin and soft tissue infections, as well as serious infections including meningitis, ventriculoperitoneal shunt infection, spondylodiscitis, prosthetic joint infection, catheter-related bacteremia, endocarditis, and pacemaker-related infections [5, 6]. S. lugdunensis is a coagulase-negative Staphylococcus with a pathogenicity and virulence more similar to Staphylococcus aureus than to other coagulase-negative Staphylococcus spp.. In fact, these organisms are frequently misidentified as S. aureus because of their morphologic appearance with yellow pigmentation and complete hemolysis when cultured on blood agar. Even so, appropriate identification of S. lugdunensis is imperative as misidentification may result in inadequately treated, prolonged, and persistent infection [6].

Indeed, S. lugdunensis comprises a niche that is distinctly different from those of S. aureus [7, 8]; S. lugdunensis is an integral part of the normal skin flora, predominant at the abdomen, groin, and lower extremities (especially the nail bed of the first toe) and is only rarely obtained from cultures of nasal swabs—a pattern of distribution distinct from S. aureus [7, 9]. Moreover, its characteristic antibiotic susceptibility profile is unusual for both coagulase-negative and coagulase-positive staphylococci. S. lugdunensis typically lacks the mecA gene, which codes for penicillin binding protein (PBP) 2a and induces methicillin resistance [10]. Moreover, because only about 20% of isolates are beta-lactamasepositive, it is often possible to use penicillin for treatment of infections caused by these organisms. In addition, S. lugdunensis is also usually susceptible to gentamicin, rifampicin, vancomycin, oxacillin, and cefoxitin, and <10% of isolates are found to be resistant to clindamycin or fusidic acid [5].

According to a 5-year consecutive record review by Herchline and Ayers, approximately 10.1% of isolated staphylococcal species that were not S. aureus or S. epidermidis were indeed identified as S. lugdunensis [11]. The most common clinical diagnoses were skin and skin structure infections (55.4%) and blood and vascular catheter infections (17.4%). For 40% of the reviewed specimens, S. lugdunensis was the sole agent isolated, and for 60% of specimens, S. lugdunensis was isolated as part of mixed flora. In only 15.4% of clinically reviewed specimens was S. lugdunensis clearly a culture contaminant or colonizing organism. The pattern of human infection identified in that study emphasizes the predominance of S. lugdunensis to affect skin and soft tissue structures [11]. Even so, this here is the first report of an association of S. lugdunensis with the fulminant manifestation of necrotizing fasciitis.

#### 4. Conclusion

S. lugdunensis is a common cause of skin and softtissue infections (SSTIs) that is likely underrated by many laboratories. S. lugdunensis is distinct in both niche, clinical spectrum of disease, and antibiotic sensitivity in comparison to other staphylococcal species. This case report demonstrates the potential of S. lugdunensis to cause SSTI as extensive as necrotizing fasciitis. As such, S. lugdunensis should be accepted

as a clinically significant human pathogen and should be distinctly identified and included in all routine bacteriological examinations. Clinicians should be acquainted with the name and the pathology of the bacterium. Moreover, this case report also identifies Group B *Streptococcus* as a synergistic microbe in this case of NF associated with *S. lugdunensis*.

#### References

- [1] C. H. Wong, H. C. Chang, S. Pasupathy, L. W. Khin, J. L. Tan, and C. O. Low, "Necrotizing fasciitis: clinical presentation, microbiology, and determinants of mortality," *Journal of Bone and Joint Surgery—Series A*, vol. 85, no. 8, pp. 1454–1460, 2003.
- [2] T. Shimizu and Y. Tokuda, "Necrotizing fasciitis," *Internal Medicine*, vol. 49, no. 12, pp. 1051–1057, 2010.
- [3] D. A. Anaya and E. P. Dellinger, "Necrotizing soft-tissue infection: diagnosis and management," *Clinical Infectious Diseases*, vol. 44, no. 5, pp. 705–710, 2007.
- [4] C. H. Wong, A. Kurup, and K. C. Tan, "Group B Streptococcus necrotizing fasciitis: an emerging disease?" European Journal of Clinical Microbiology and Infectious Diseases, vol. 23, no. 7, pp. 573–575, 2004.
- [5] C. Hellbacher, E. Tornqvist, and B. Soderquist, "Staphylococcus lugdunensis: clinical spectrum, antibiotic susceptibility, and phenotypic and genotypic patterns of 39 isolates," Clinical Microbiology and Infection, vol. 12, no. 1, pp. 43–49, 2006.
- [6] H. Seifert, D. Oltmanns, K. Becker, H. Wisplinghoff, and C. Von Eiff, "Staphylococcus lugdunensis pacemaker-related infection," Emerging Infectious Diseases, vol. 11, no. 8, pp. 1283–1286, 2005.
- [7] L. Bieber and G. Kahlmeter, "Staphylococcus lugdunensis in several niches of the normal skin flora," Clinical Microbiology and Infection, vol. 16, no. 4, pp. 385–388, 2010.
- [8] T. Y. Tan, S. Y. Ng, and J. He, "Microbiological characteristics, presumptive identification, and antibiotic susceptibilities of *Staphylococcus lugdunensis*," *Journal of Clinical Microbiology*, vol. 46, no. 7, pp. 2393–2395, 2008.
- [9] S. Bocher, B. Tonning, R. L. Skov, and J. Prag, "Staphylococcus lugdunensis, a common cause of skin and soft tissue infections in the community," Journal of Clinical Microbiology, vol. 47, no. 4, pp. 946–950, 2009.
- [10] J. R. Ebright, N. Penugonda, and W. Brown, "Clinical experience with *Staphylococcus lugdunensis* bacteremia: a retrospective analysis," *Diagnostic Microbiology and Infectious Disease*, vol. 48, no. 1, pp. 17–21, 2004.
- [11] T. E. Herchline and L. W. Ayers, "Occurrence of *Staphylococcus lugdunensis* in consecutive clinical cultures and relationship of isolation to infection," *Journal of Clinical Microbiology*, vol. 29, no. 3, pp. 419–421, 1991.